

# Systems Biology

Non-linear signal processing and hormetic signal-response rates in the TOR pathway ?

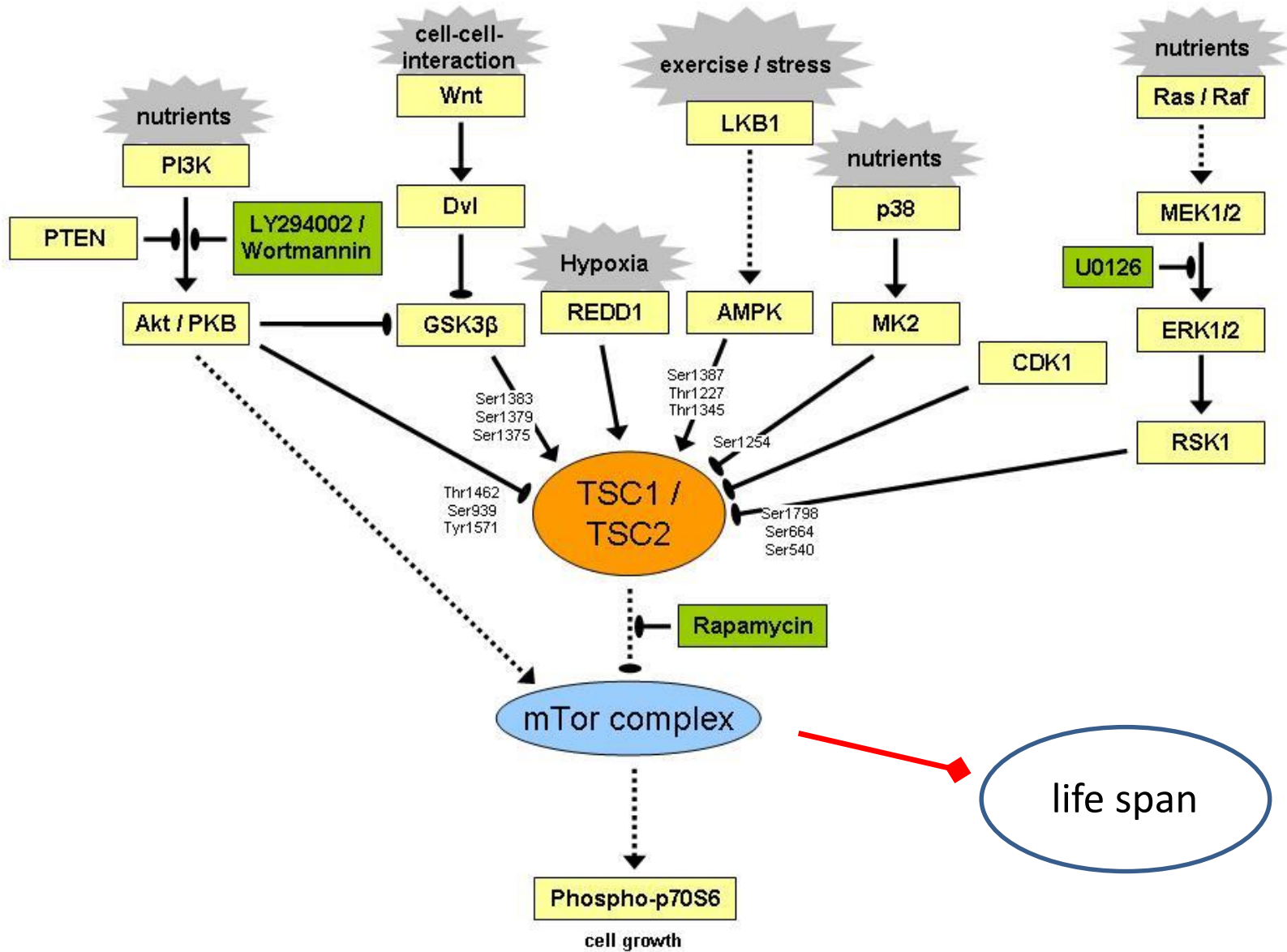
Uwe Menzel, 2015

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# Target Of Rapamycin (TOR) pathway

- TOR plays a crucial role for a number of cellular processes such as ageing [Johnson 2013, Evans 2010, Lamming 2011, Bonawitz 2007], tumour formation [Populo 2012], adipogenesis, formation of insulin resistance, and activation of the immune response [Thomson 2009].
- Decreased TOR activity has been found to slow aging in *S. cerevisiae*, *C. elegans*, and *D. melanogaster*. The mTOR inhibitor rapamycin has been confirmed to increase lifespan in mice by independent groups ....
  - this can, for instance, be accomplished by activation of the proteins TSC1/2 (tuberous sclerosis complex 1+2)
- central role for regulation of cellular adaption and homeostasis
- responds to a large number of intracellular and extracellular signals
- regulates metabolism, growth, proliferation, ...



# Hormesis, Hormetic Response

- describes effects of an agent on a cell, an organism etc.
  - beneficial effect after exposure to low doses
  - toxic – or even lethal - effect for higher doses of the same agent.
- “For every substance, small doses stimulate, ..., large doses kill.”  
([Arndt–Schulz rule](#))
- **more general:** Hormesis is an adaptive response of biological systems to moderate (transient) levels of stress factors

# Stress factor: Reactive Oxygen Species (ROS)

- **ROS** (“free radicals”) is a main stress factor
- “free radical”: atom, molecule, or ion with an unpaired valence electron
  - superoxide (  $O_2^-$  )
  - hydrogen peroxide ( $H_2O_2$ )
  - peroxyntirite ( $OOONO^-$ )
- Consequences:
  - cancer
  - oxidation of LDL → plaque in arteries → heart disease, stroke
  - cross-linking between fat and protein molecules

# “Traditional” free-radical theory of aging

- D. Harman, 1956: [Free radical theory of aging](#)
- “free radicals” produce cumulative damage of cells and shorten lifespan
- drugs ([antioxydants](#)) act against free radicals
  - $\beta$ -carotene, superoxide dismutase, vitamins A, C, E, coenzyme Q [ubiquinol], glutathione, curcumin [E100]

# New findings



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Contents lists available at ScienceDirect

## Experimental Gerontology

journal homepage: [www.elsevier.com/locate/expgero](http://www.elsevier.com/locate/expgero)

Review

How increased oxidative stress promotes longevity and metabolic health:  
The concept of mitochondrial hormesis (mitohormesis)

Michael Ristow<sup>a,b,\*</sup>, Kim Zarse<sup>a</sup>

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## New findings

# Antioxidants prevent health-promoting effects of physical exercise in humans

Michael Ristow<sup>a,b,1,2</sup>, Kim Zarse<sup>a,2</sup>, Andreas Oberbach<sup>c,2</sup>, Nora Klötting<sup>c</sup>, Marc Birringer<sup>a</sup>, Michael Kiehntopf<sup>d</sup>, Michael Stumvoll<sup>c</sup>, C. Ronald Kahn<sup>e</sup>, and Matthias Blüher<sup>c,2</sup>

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Contributed by C. Ronald Kahn, March 31, 2009 (sent for review March 14, 2009)



# High oxidative damage levels in the longest-living rodent, the naked mole-rat

Blazej Andziak,<sup>1</sup> Timothy P. O'Connor,<sup>2</sup> Wenbo Qi,<sup>3</sup>  
Eric M. DeWaal,<sup>4</sup> Anson Pierce,<sup>3,5</sup>

We compare antioxidant defenses (reduced glutathione, GSH), redox status (GSH/GSSG), as well as lipid (malondialdehyde and isoprostanes), DNA (8-OHdG), and protein (carbonyls) oxidation levels in urine and various tissues from both mole-rats and similar-sized mice. Significantly lower GSH and GSH/GSSG in mole-rats indicate poorer antioxidant capacity and a surprisingly more pro-oxidative cellular environment, manifested by 10-fold higher levels of *in vivo* lipid peroxidation. Furthermore, mole-rats exhibit greater levels of accrued oxidative damage to lipids (twofold), DNA (~two to eight times) and proteins (1.5 to 2-fold) than physiologically age-matched mice, and equal to that of same-aged mice.

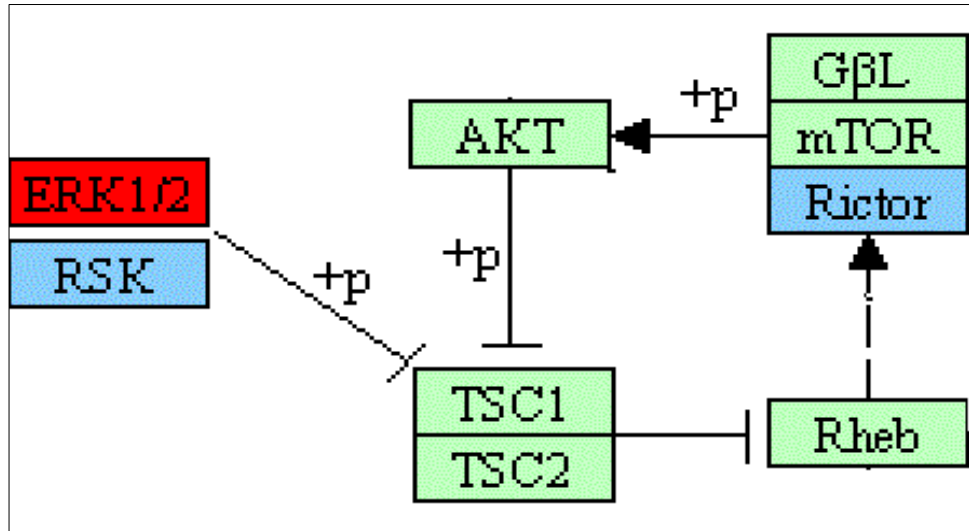
# Mathematical model of hormesis in the TOR pathway

- **Goal:** model the following qualitative behavior mathematically:
  - short or low-intensity **pulses** of ROS → activation of a “**defense enzyme**” (against ROS)
  - long or high-intensity pulses → defense loop cannot be sustained, “**destructive molecule**” is released

“For every substance, small doses stimulate, moderate doses inhibit, large doses kill.”

Arndt-Schulz rule

## TOR contains a double-negative feedback loop (?)



### TOR-C2 complex

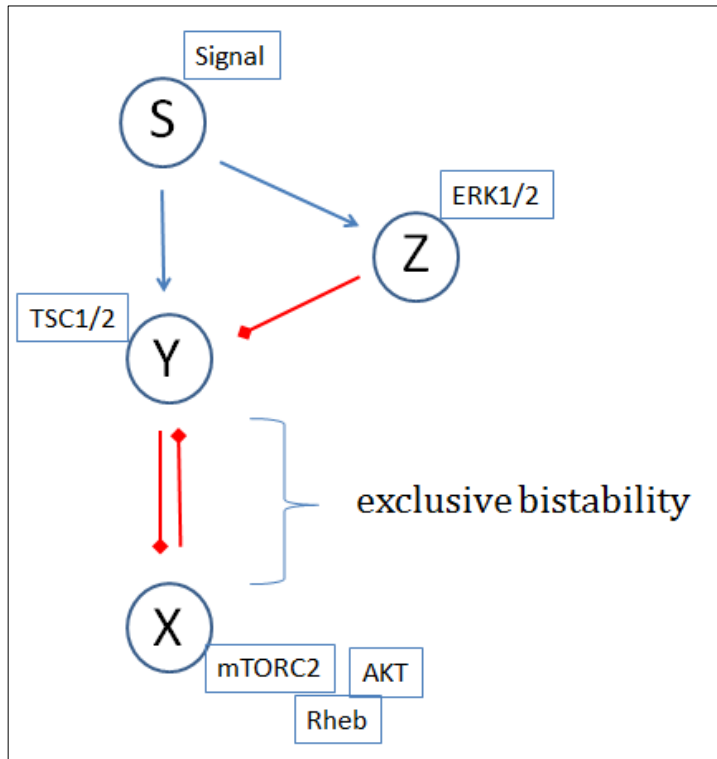
Arrow-headed: activation  
T-shaped: inhibition

KEGG pathway

The double negative loop forms **exclusive bistability** between

- TOR-C2 complex (GβL, mTOR, Rictor) – “**destructive**”
- and TSC1/2 – “**defense enzyme**”

**Note:** It is not entirely sure if the KEGG pathway data are correct.



## Schematic plot of the double-negative feedback loop

- AKT, TOR, and Rheb are merged into one variable **X**
- **X** suppresses the TSC1/2 complex (**Y**).
- TSC1/2 in turn suppresses **X** → exclusive bistability (red lines)
- Also ERK1/2 (**Z**) suppresses TSC1/2
- ERK1/2 and TSC1/2 are triggered by an external signal **S** (ROS)

# Nonlinear system of ODE

$$\frac{dx}{dt} = \alpha_x - \beta_x \cdot x - \beta_{xy} \cdot h(y, K_{xy}) \cdot x \quad \boxed{\text{mTOR-C2}}$$

$$\frac{dy}{dt} = I_y \cdot S(t) + \alpha_y - \beta_y \cdot y - \beta_{xy} \cdot h(x, K_{yx}) \cdot y - \beta_{yz} \cdot h(z, K_{yz}) \cdot y \quad \boxed{\text{TSC1/2}}$$

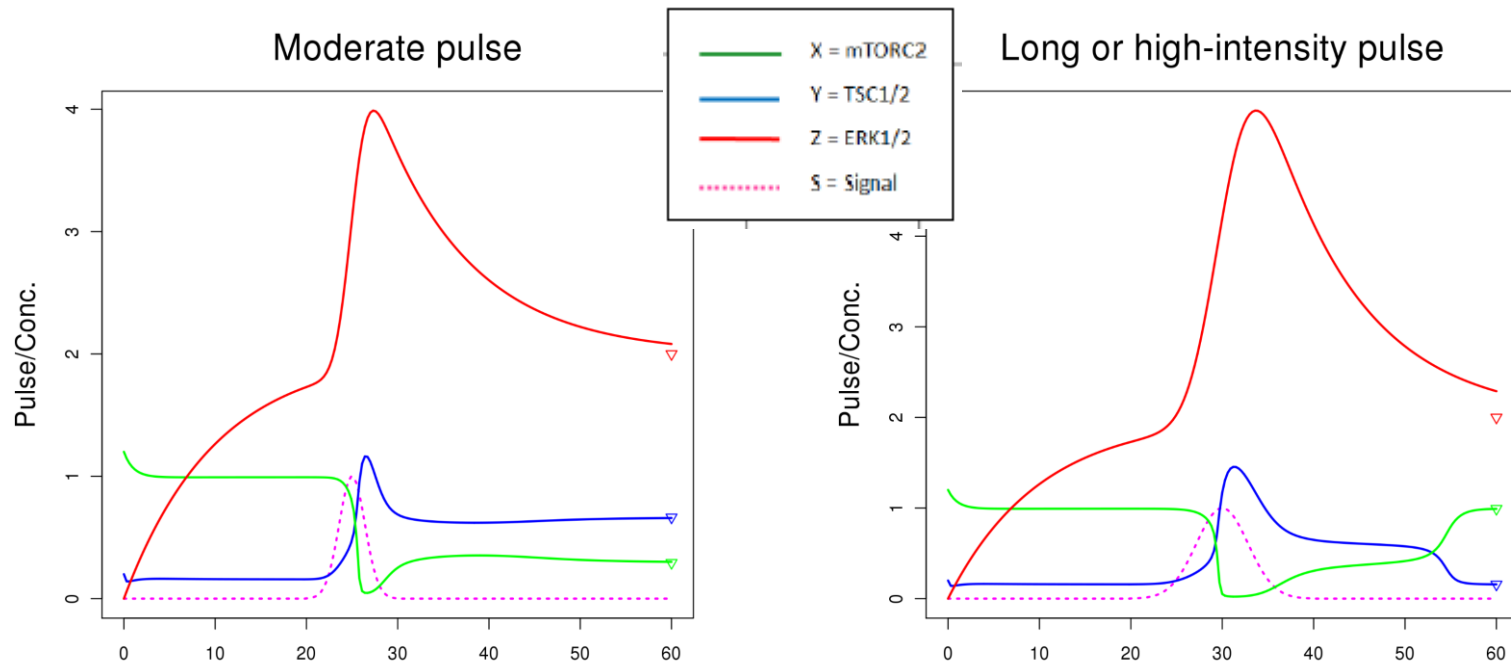
$$\frac{dz}{dt} = I_z \cdot S(t) + \alpha_z - \beta_z \cdot z \quad \boxed{\text{ERK1/2}}$$

$$h(x, K) = \frac{\left(\frac{x}{K}\right)^4}{1 + \left(\frac{x}{K}\right)^4} \quad \text{Hill term}$$

- $\alpha_i$  : basic time-independent synthesis terms
- $\beta_i$ : spontaneous degradation of enzyme  $i$

$\boxed{\text{R: deSolve (LSODA), nleqslv, ...}}$

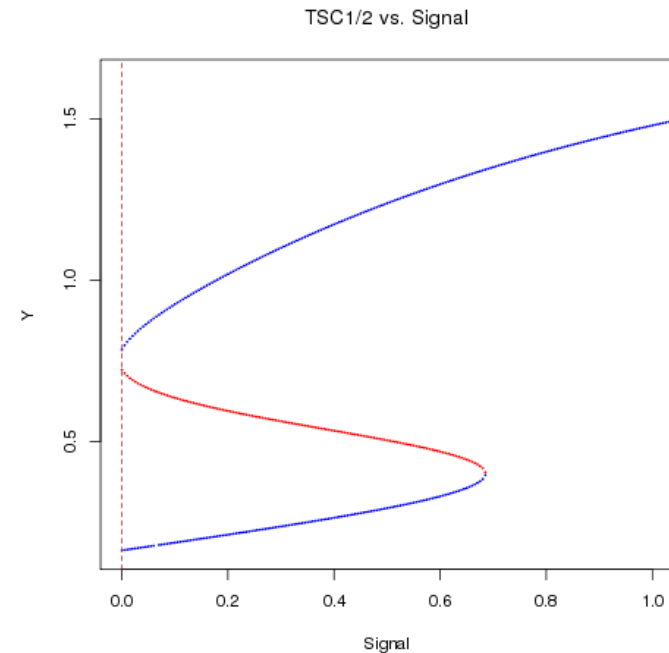
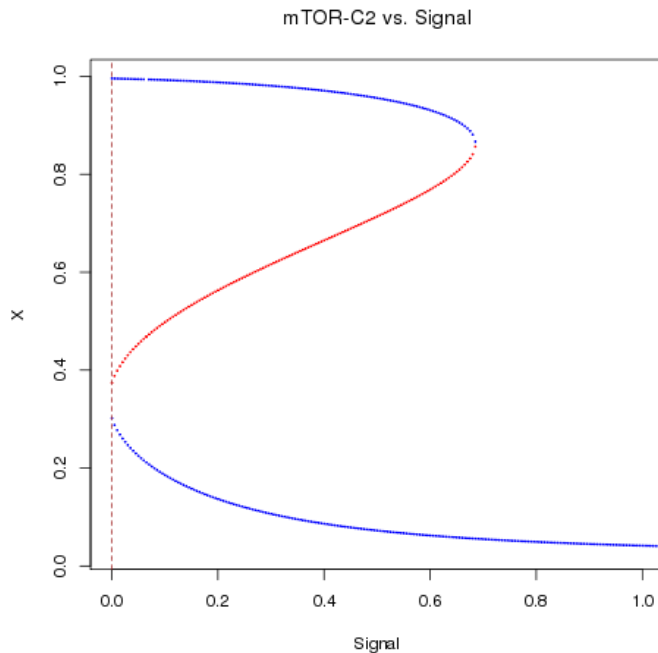
# Results of the calculations



- external signal (dotted), TOR-C2, TSC1/2, ERK1/2
- left: ROS signal with half-width 5
- right: ROS signal with half-width 10 (same amplitude)
- (stable) steady state values: triangles, same colours
- moderate signal: defense loop (TSC1/2) permanently upregulated
- strong signal: defense loop cannot be maintained
- → characteristics of hormesis

# Bistability

- above:  $X, Y$  have 2 different stable states for signal  $S = 0 \rightarrow$  bifurcation
- exhaustive search algorithm to find all solutions for different  $S(t) = S$



- TOR-C2 (left) and TSC1/2 (right) display irreversible hysteresis
- **blue lines:** stable states ; **red lines:** unstable states

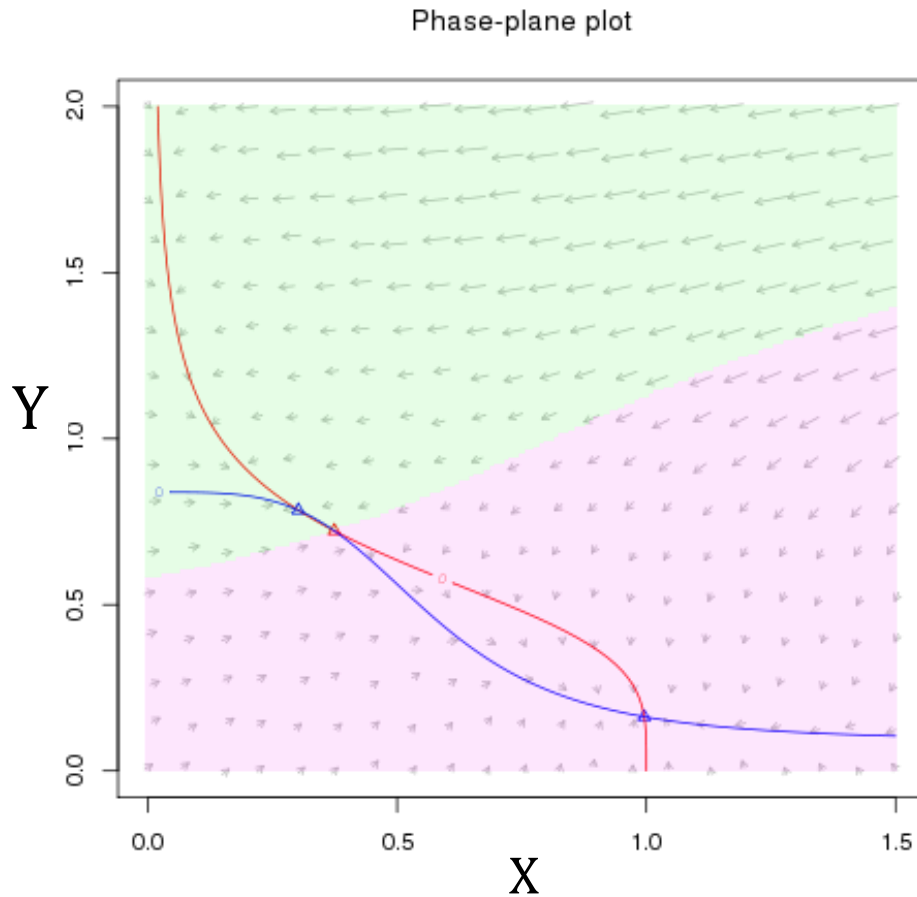
# Why does the defense state vanish for strong ROS signals ?

- irreversible hysteresis explains that ROS signal brings the system into defense state (TSC1/2 up, TOR-C2 down)
- ... **but cannot explain why strong pulses destroy the defense state**
- the latter is a consequence of non-stationarity of ERK1/2 (variable  $Z$ )
- this can be understood by looking at the phase portraits for different values of  $Z$





# Phase portrait for equilibrium state of ERK1/2 (Z)



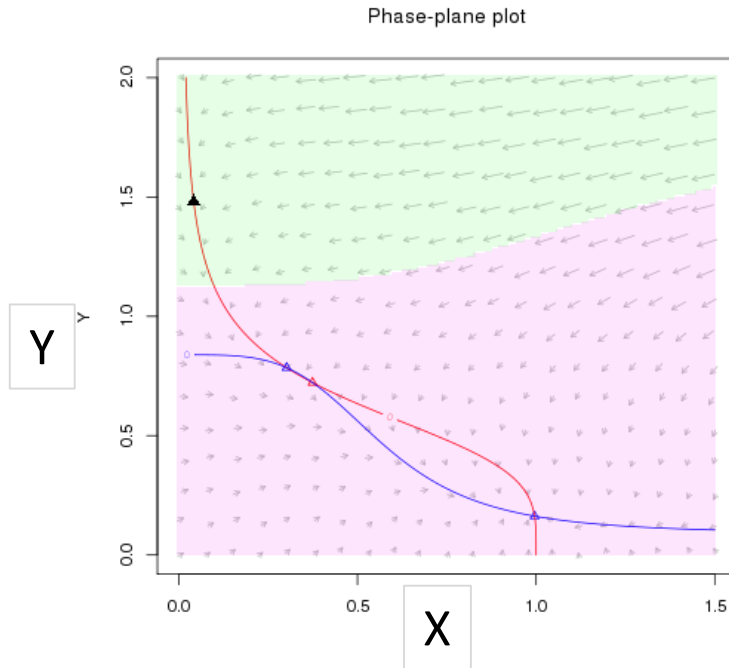
$$Z_0 = \frac{\alpha_z + I_z \cdot S_0}{\beta_z}$$

- steady state for ODE
- two stable solutions
- one instable solution
- nullclines
- attraction domains

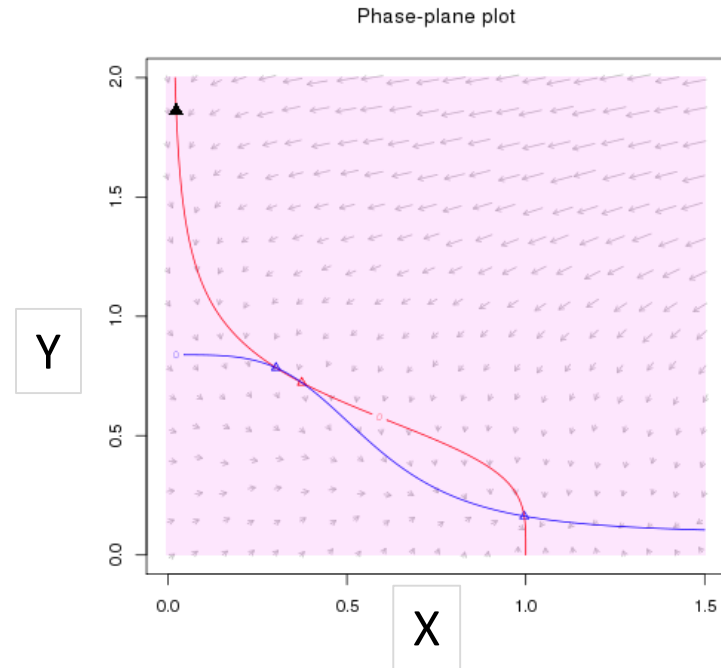
# Phase portraits for non-equilibrium states of ERK1/2 (Z)

- situation described above does not apply to times short after signal termination
- .... where ERK1/2 (Z) is **not in equilibrium**
- system "**overheated**" shortly after exposure to ROS

# Phase portraits for non-equilibrium states of ERK1/2 (Z)



$(X, Y, Z) = (0.0416, 1.48, 2.0)$   
(black triangle)



$(X, Y, Z) = (0.0227, 1.864, 3.99)$   
(black triangle)

right plot: system state after signal in attractor region of lower equilibrium point → system returns to this unfavourable state

Response of the three-gene network motif to (ROS-) pulses of varying duration or intensity:

Pulse strength	Destructive mol.	Defense enzyme
moderate	Off	On
long	On	Off

## Back to the hormesis model:

- On a so far simple level, the model meets our expectations:
  - moderate pulses bring up a “defense enzyme” (dismutase-associated?, peroxydase-associated?).
  - At long pulse durations, this enzyme can no longer be sustained
  - over a wide range of parameters, we find the same qualitative behavior (luckily ...)
- Critics:
  - it can't be that simple
  - if just a third gene (“destructive molecule”) would bring down the defense enzyme again, evolution would have knocked it out, **would it?**

## Progress:

- Computations work (for practically arbitrary network motifs), large number of genes doesn't seem to be a (computational) problem
- Qualitative differences in molecule levels in response to varying signal durations modelled
- Qualitative results similar for a large range of parameter values
- Stable levels of a “defense enzyme” can be set up at appropriate pulse levels (modeling of varying pulse height yields similar behavior)
- can be expanded to more genes/equations and may lead to a model for a known pathway:

## To think about:

- Detailed analysis of the reactions in the mTOR pathway
- Treat transcript- and protein concentrations as separate variables (e.g. Michaelis-Menten kinetics for phosphorylation) ?
- Include active/inactive states of transcription factors?
- How to compare with experimental results? (compare measured  $\leftrightarrow$  calculated ratios at steady state)
- Modeling of the source term in the equations
- Fit to RNA-Seq data (e.g. MC, Bayes, simulated annealing – seems feasible)

# Appendix

Non-linear signal processing and hormetic signal-response rates in the TOR pathway ?

Uwe Menzel, 2015

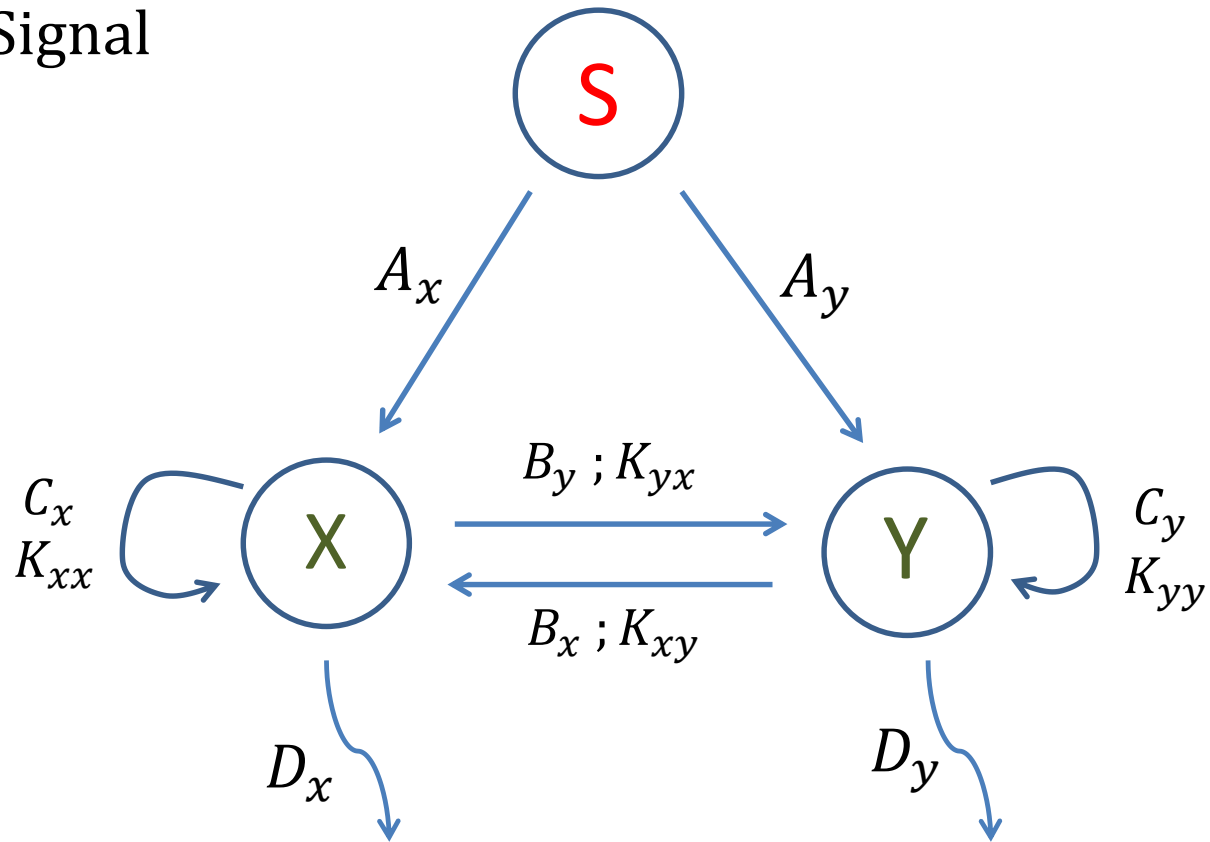
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[www.matstat.org](http://www.matstat.org)

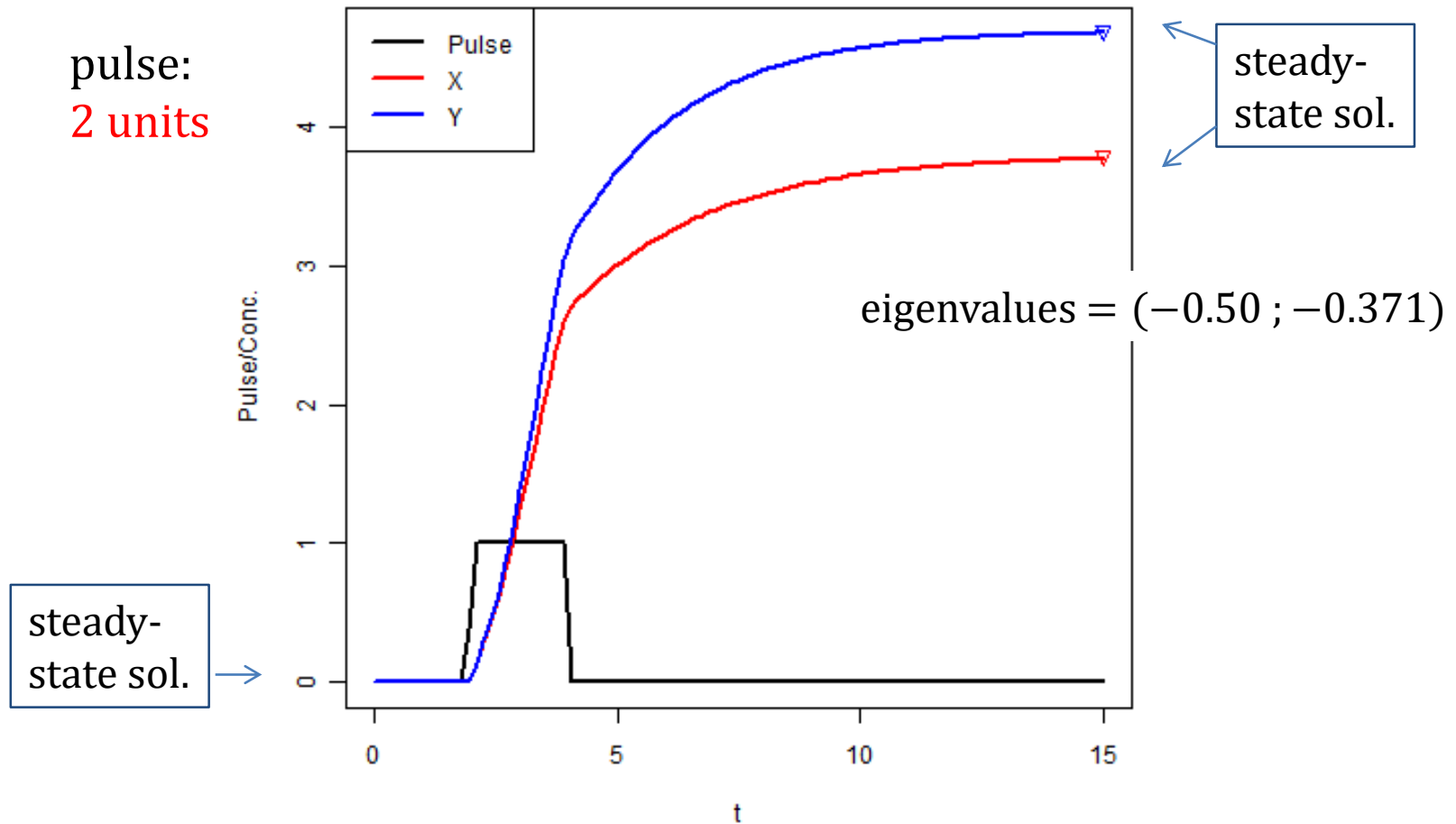


# Simple motif: double-positive regulated

**S**: Signal



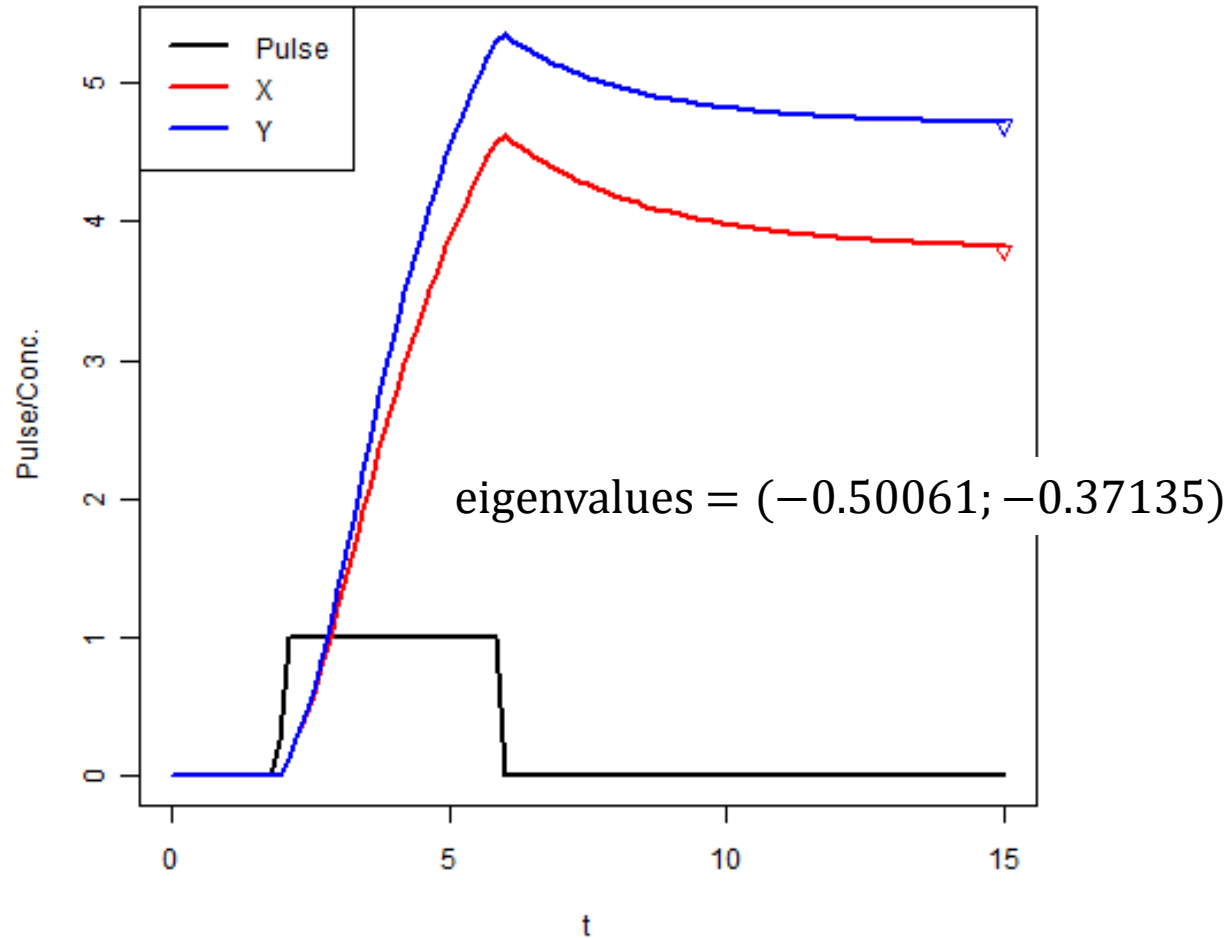
# Short-pulse response



- both genes remain ON even after the pulse has terminated, i.e. defense enzyme (blue) is established
- concentrations increase even after pulse termination – approach “upper” steady state

## Long-pulse response

pulse:  
4 units



- **longer pulse:** both concentrations obtain a higher level during the pulse but approach the **same** steady-state value after pulse termination

# Least Squares Fit with Non-linear Optimization

$$0 = \alpha_x - \beta_x \cdot x - \beta_{xy} \cdot h(y, K_{xy}) \cdot x$$

$$0 = \alpha_y - \beta_y \cdot y - \beta_{xy} \cdot h(x, K_{yx}) \cdot y - \beta_{yz} \cdot h(z, K_{yz}) \cdot y$$

$$0 = \alpha_z - \beta_z \cdot z$$



optim  
constrOptim

Least squares: Minimize Sum of Squared Errors (SSE)

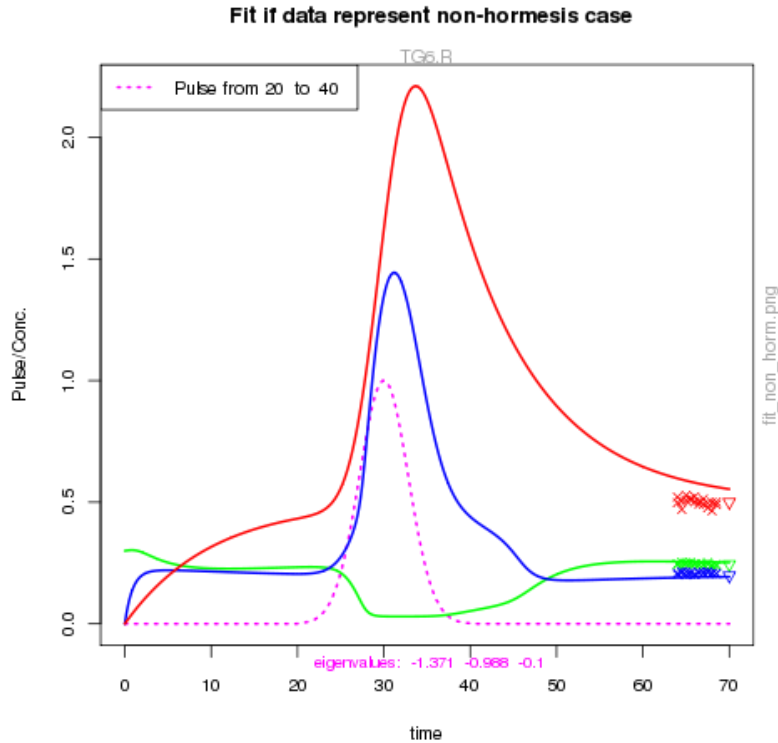
$$SSE(A_1, A_2, A_3) = \sum_{i=1}^N A_i^2 + B_i^2 + C_i^2 \quad \Rightarrow \quad \text{Minimum}$$

$$A_i = \alpha_x - \beta_x \cdot x_i - \beta_{xy} \cdot h(y_i, K_{xy}) \cdot x_i$$

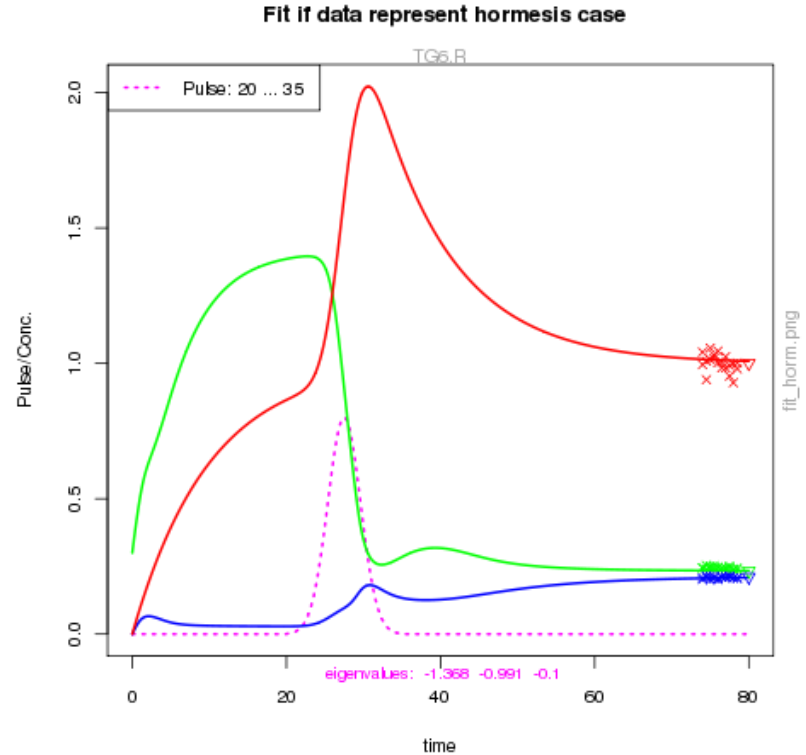
$$B_i = \alpha_y - \beta_y \cdot y_i - \beta_{xy} \cdot h(x_i, K_{yx}) \cdot y_i - \beta_{yz} \cdot h(z_i, K_{yz}) \cdot y_i$$

$$C_i = \alpha_z - \beta_z \cdot z_i$$

# Fit to experimental data

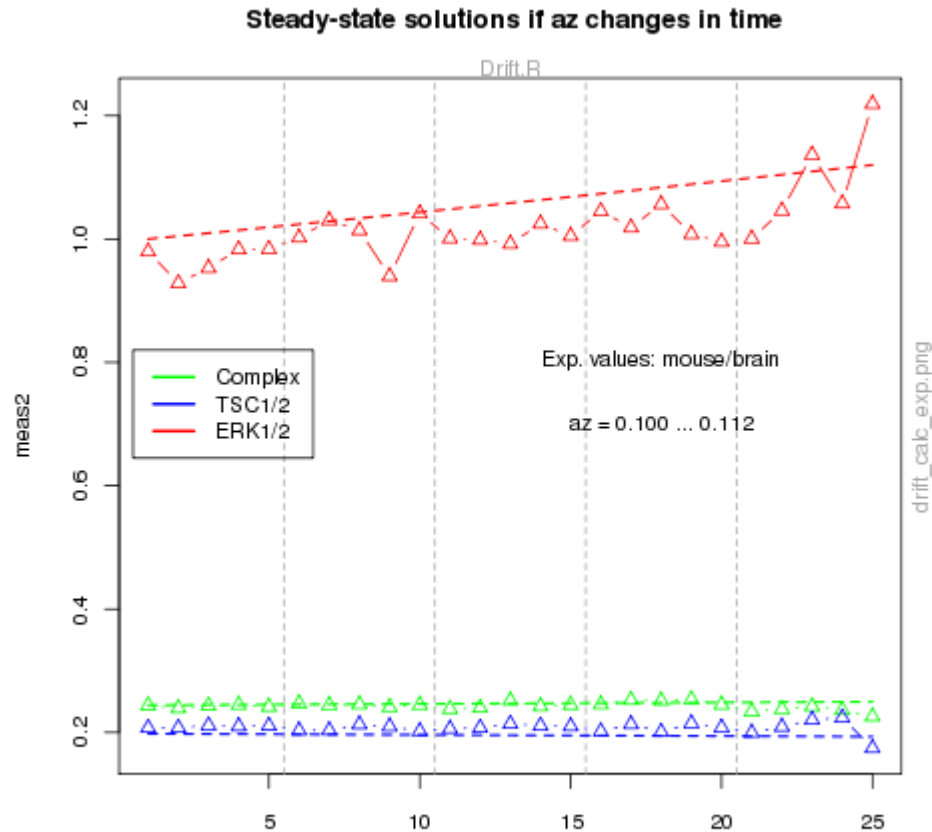


... if the data represent state 1



... if the data represent state 2  
("defense loop")

# Long-term behavior (ageing)



Increase of ERK1/2 and stationarity of both TSC1/2 and the “complex” can (only) be explained by an increasing intrinsic production rate of ERK1/2 ( $\alpha_z$ ).